# Tropical Journal of Phytochemistry & Pharmaceutical Sciences

Available online at <a href="https://www.tjpps.org">https://www.tjpps.org</a>

# Original Research Article

# In vitro Cytotoxicity, Antioxidant, and Anti-inflammatory Activities of Eucalyptus camaldulensis Leaf Extracts

Ibrahim A. Usman<sup>1\*</sup>, and Maryam M. Ali<sup>1</sup>

<sup>1</sup>Department of Pure and Industrial Chemistry, Faculty of Physical Sciences, Bayero University, Kano, Nigeria.

# ABSTRACT

Chronic inflammation and oxidative stress are key contributors to the onset and progression of several degenerative diseases. Medicinal plants play a vital role in their management due to their rich content of bioactive compounds, which possess antioxidant and anti-inflammatory properties. This study evaluated the cytotoxic, antioxidant, and anti-inflammatory activities of *Eucalyptus camaldulensis* leaf extracts collected in Kano, Nigeria. The powdered leaves were extracted with ethanol and fractionated into n-hexane, chloroform, ethyl acetate, and methanol fractions. Antioxidant activity was assessed using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay, anti-inflammatory activity by inhibition of protein denaturation, and cytotoxicity using the brine shrimp lethality test. The methanol extract exhibited the strongest antioxidant effect (IC<sub>50</sub> = 4.785  $\mu$ g/mL), which was lower in potency than ascorbic acid (IC<sub>50</sub> = 0.943  $\mu$ g/mL). The n-hexane fraction demonstrated the highest anti-inflammatory activity (IC<sub>50</sub> = 3.215  $\mu$ g/mL) and cytotoxicity (LC<sub>50</sub> = 3.810  $\mu$ g/mL), while the methanol fraction was non-toxic (LC<sub>50</sub> > 1000  $\mu$ g/mL). These findings indicate that *Eucalyptus camaldulensis* possesses solvent-dependent biological activities, with the methanol fraction showing strong antioxidant and safe pharmacological properties, and the n-hexane fraction displaying significant anti-inflammatory and cytotoxic effects. The study provides scientific support for the traditional use of *Eucalyptus camaldulensis* and suggests its potential as a source of natural therapeutic agents. Further studies should focus on the isolation, characterization, and *in vivo* evaluation of its bioactive compounds.

Keywords: Eucalyptus camaldulensis, antioxidant Activity, anti-Inflammatory Activity, aytotoxicity, DPPH assay, protein denaturation, brine shrimp lethality.

Received 02 October 2025 Revised 29 October 2025 Accepted 30 October 2025 Published online 01 November 2025 **Copyright:** © 2025 Usman *et al.* This is an open-access article distributed under the terms of the <u>Creative Commons</u> Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

# Introduction

Medicinal plants are recognized as important sources of bioactive compounds with therapeutic potential<sup>1, 2</sup>. Chronic inflammation and oxidative stress are key contributors to the onset and progression of various diseases, including cancer, diabetes, and cardiovascular disorders3. Consequently, research on medicinal plants as natural antioxidants and anti-inflammatory agents has intensified, aiming to identify safer and more effective alternatives to synthetic drugs4. Eucalyptus camaldulensis (river red gum), a member of the Myrtaceae family, is widely distributed and frequently used in traditional medicine<sup>3</sup>. Previous studies have documented its antimicrobial<sup>5</sup>, anti-diabetic<sup>6</sup>, and cytotoxic<sup>7</sup> properties. However, many of these investigations focused on samples from other regions or included limited biological analysis. Therefore, comprehensive validation of the antioxidant, anti-inflammatory, and cytotoxic properties of Nigerian populations of E. camaldulensis is lacking, particularly considering possible phytochemical variations due to environmental and climatic differences<sup>8,9</sup>. The present study was therefore designed to evaluate the cytotoxic, antioxidant, and antiinflammatory activities of different solvent extracts of Eucalyptus camaldulensis leaves collected in Kano, Nigeria. This study not only aims to provide scientific justification for its ethnomedicinal use but also to explore its potential as a source of bioactive compounds for pharmaceutical development.

\*Corresponding author. mail: <u>iau1900662.che@buk.edu.ng</u> Tel: +234-8060763763

Citation: Usman I.A., Ali M.M. *In vitro* cytotoxicity, antioxidant, and anti-inflammatory activities of *Eucalyptus camaldulensis* leaf extracts. Trop J Phytochem Pharm Sci, 2025; 4(10): 404 – 407 <a href="http://www.doi.org/10.26538/tjpps/v4i10.5">http://www.doi.org/10.26538/tjpps/v4i10.5</a>

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

Medicinal plants are recognized as important sources of bioactive compounds with therapeutic potential<sup>1, 2</sup>. Chronic inflammation and oxidative stress are key contributors to the onset and progression of various diseases, including cancer, diabetes, and cardiovascular disorders3. Consequently, research on medicinal plants as natural antioxidants and anti-inflammatory agents has intensified, aiming to identify safer and more effective alternatives to synthetic drugs4. Eucalyptus camaldulensis (river red gum), a member of the Myrtaceae family, is widely distributed and frequently used in traditional medicine<sup>3</sup>. Previous studies have documented its antimicrobial<sup>5</sup>, antidiabetic6, and cytotoxic7 properties. However, many of these investigations focused on samples from other regions or included limited biological analysis. Therefore, comprehensive validation of the antioxidant, anti-inflammatory, and cytotoxic properties of Nigerian populations of E. camaldulensis is lacking, particularly considering possible phytochemical variations due to environmental and climatic differences<sup>8,9</sup>. The present study was therefore designed to evaluate the cytotoxic, antioxidant, and anti-inflammatory activities of different solvent extracts of Eucalyptus camaldulensis leaves collected in Kano, Nigeria. This study not only aims to provide scientific justification for its ethnomedicinal use but also to explore its potential as a source of bioactive compounds for pharmaceutical development.





Figure 1: Eucalyptus Camaldulensis Tree (Left) and Leaves (Right)

#### **Materials and Methods**

# Plant Collection and Authentication

Fresh leaves of Eucalyptus camaldulensis were collected in December 2024. The plant was identified and authenticated by a taxonomist in the Department of Biological Sciences, Bayero University, Kano. A voucher specimen (No. BUKHAN 0347) was deposited at the Bayero University Herbarium for future reference

#### Extraction and Fractionation

The powdered leaves (50 g) were macerated in 200 mL of 95% ethanol for seven days with intermittent shaking. Afterward, the mixture was decanted and filtered using Whatman filter paper to separate the liquid extract from the solid residue. The filtrate was left to evaporate at room temperature for three days to obtain the crude extract, which yielded 6.4 g. One gram (1 g) was removed for crude analysis, while the remaining 5.4 g was fractionated successively with solvents of increasing polarity (n-hexane, chloroform, ethyl acetate, and methanol), following the procedure described by Usman et al. (2021)<sup>10</sup>.

# Antioxidant Assay (DPPH Radical Scavenging Activity)

The antioxidant activity of the extracts was evaluated using the DPPH radical scavenging method as described by Gülcin (2009)<sup>11</sup> and Falodun et al.  $(2025)^{12}$  with slight modifications. Briefly, different concentrations of the extracts (7.81-1000  $\mu$ g/mL) were mixed with DPPH solution in methanol and incubated in the dark at room temperature for 30 minutes. Absorbance was measured at 517 nm using a UV-visible spectrophotometer, with ascorbic acid serving as the positive control. IC50 values were determined from dose–response data using SPSS software.

# Anti-Inflammatory Assay (Protein Denaturation Method)

The anti-inflammatory activity of the extracts was assessed using the *in vitro* protein denaturation assay. 20  $\mu$ L of each extract (7.81-1000  $\mu$ g/mL) was mixed with 0.2 mL of fresh egg albumin and 180  $\mu$ L of phosphate-buffered saline (PBS) in a 96-well microplate. The mixture was incubated in an oven at 70 °C for 15 minutes, allowed to cool, and the turbidity was measured at 660 nm. IC<sub>50</sub> values were calculated from dose-response data using SPSS software.

# Cytotoxicity Assay (Brine Shrimp Lethality Test)

The cytotoxic activity of the extracts was evaluated using the brine shrimp lethality test as described by Meyer et al.  $(1982)^{13}$  and further applied in similar studies by Amuamuta  $(2023)^{14}$  with slight modifications.  $Artemia\ salina\ (brine\ shrimp\ larvae)$  was used as the test organism. The eggs were hatched in a 1000 mL beaker containing seawater and kept under constant illumination and aeration for  $24-72\ h$  until the nauplii (larvae) emerged. Stock solutions of the extracts  $(10,000\ \mu g/mL)$  were prepared by dissolving 30 mg of each extract in 3 mL of ethanol. Working concentrations of 1000, 100, and 10  $\mu g/mL$ 

were prepared in test tubes in triplicate. After adding 10 active nauplii to each tube, the total volume was adjusted to 5 ml of seawater. After 24 h of exposure, the number of surviving nauplii was counted, and the percentage mortality was calculated. LC<sub>50</sub> values were determined from the mortality data using probit regression analysis in SPSS software.

# **Results and Discussion**

#### Extraction Yield

The crude ethanol extracts of *Eucalyptus camaldulensis* leaves yielded 6.4 g of residue. Fractionation produced n-hexane (0.23 g), chloroform (0.86 g), ethyl acetate (1.02 g), and methanol (1.56 g) fractions. The highest yield was obtained from methanol, consistent with its ability to extract polar phytochemicals such as flavonoids and phenolics<sup>5</sup>. This agrees with previous studies that reported higher polar extract yields in *Eucalyptus* species<sup>15</sup>.

#### Antioxidant Activity

The DPPH radical scavenging activity showed that the methanol fraction had the strongest activity (ICso = 4.785  $\mu$ g/mL), followed by the crude ethanol extract (ICso = 15.854  $\mu$ g/mL), while ascorbic acid, used as a positive control, had ICso = 0.943  $\mu$ g/mL (Table 2, Figure 2).

**Table 1:** Extraction yield of fractions

Fraction	Weight (g)	Characteristics
n-Hexane	0.23	Oily, dark green
Chloroform	0.86	Viscous, dark green
Ethyl acetate	1.02	Clumpy, brownish green
Methanol	1.56	Powdery, yellowish brown

This indicates that the methanol fraction is rich in polyphenolic compounds capable of donating hydrogen atoms to neutralize free radicals. Previous studies have shown similar antioxidant properties of *Eucalyptus camaldulensis* attributed to flavonoids and tannins<sup>9, 15, 16</sup>. The results justify its ethnomedicinal use as an antioxidant agent.

# Anti-Inflammatory Activity

The protein denaturation assay revealed that the n-hexane fraction demonstrated the strongest activity (caused the least protein denaturation, indicating higher membrane stabilization potential,  $IC_{50}=3.215~\mu g/mL)$ , followed by the methanol fraction ( $IC_{50}=6.713~\mu g/mL)$  (Table 3, Figure 3). The high activity of the nonpolar fraction suggests that lipophilic compounds, possibly terpenoids and fatty acid derivatives, play a significant role in membrane stabilization and inhibition of protein denaturation. These findings are consistent with earlier reports that hydrophobic phytochemicals from Eucalyptus species exhibit significant anti-inflammatory effects  $^{6,\,8,\,9,\,17}$ .

**Table 2:** Percentage DPPH radical scavenging activity of *Eucalyptus camaldulensis* extracts compared with the standard (Ascorbic acid)

Conc. (µg/mL)	n-Hexane inhibition)	(% <b>Chloroform</b> inhibition)	(% Ethyl acetate (% inhibition)	Methanol inhibition)	(% Crude EtOH inhibition)	(% Ascorbic acid (% inhibition)
1000	93.55	94.80	97.97	96.67	111.33	91.18
500	95.55	95.46	95.02	94.65	100.16	85.21
250	85.79	93.30	93.54	93.39	95.19	78.64
125	47.47	90.31	71.06	92.27	91.75	77.58
62.5	23.81	63.89	42.83	92.09	91.47	73.71
31.25	10.02	27.91	9.81	90.19	73.98	71.73
15.625	3.59	15.04	6.25	71.65	48.96	68.13
7.8125	3.35	12.56	3.97	42.64	25.10	66.34
$IC_{50}$ (µg/mL)	113.71	45.48	76.57	4.79	15.85	0.94

# Cytotoxicity

Brine shrimp (Artemia salina) lethality results indicated that the n-hexane fraction was the most cytotoxic (LC<sub>50</sub> =  $3.810 \mu g/mL$ ), while

the chloroform and ethyl acetate fractions showed moderate toxicity. The methanol fraction was non-toxic (LCso > 1000  $\mu$ g/mL) (Table 4, Figure 4). The brine shrimp lethality test was employed as a preliminary bioassay to assess general cytotoxicity and to predict potential

anticancer activity of the plant extracts. The marked toxicity observed in the n-hexane fraction suggests the presence of bioactive lipophilic compounds with possible anticancer or antiproliferative properties. This observation is in agreement with the findings of Mohammed (2021)<sup>7</sup> and Kumar (2014)<sup>18</sup>, who reported cytotoxic effects of *Eucalyptus* extracts against cancer cell lines. Conversely, the non-toxic nature of the methanol fraction indicates its safety and supports its potential use as a natural antioxidant in nutraceutical applications <sup>19, 20, 21</sup>.

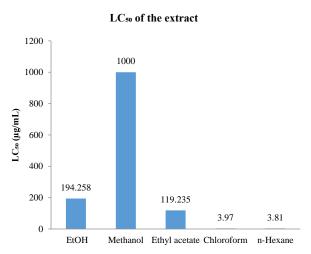
DPPH IC50 of the extract

#### 

**Figure 2:** Antioxidant activity of *Eucalyptus camaldulensis* extracts compared with ascorbic acid

#### Anti-Inflamatory IC50 of the extract 50 45.38 45 40 35 ICso (µg/mL) 30 25 20 15 9.08 8.55 6.71 10 3.22 5 0 N-Hexane Chloroform Ethyl Methanol Ethanol acetate

**Figure 3:** Anti-inflammatory activity of *Eucalyptus camaldulensis* extracts



**Figure 4:** Cytotoxicity of *Eucalyptus camaldulensis* extracts (Brine shrimp assay)

**Table 3:** Percentage inhibition of protein denaturation (anti-Inflammatory activity) of *Eucalyptus camaldulensis* leaf

		extr	acetate         (%         EtOH (% inhibition)           on)         (%         inhibition)           inhibition)         152.19         96.51         92.56           78.14         94.82         90.80           74.97         94.27         92.09           66.38         91.48         83.39           64.95         88.87         90.05		
Conc. (µg/mL)	n-Hexane (% inhibition)	Chloroform (% inhibition)	acetate (%	(%	EtOH (%
1000	94.52	91.31	152.19	96.51	92.56
500	91.98	88.42	78.14	94.82	90.80
250	84.37	84.00	74.97	94.27	92.09
125	82.11	79.67	66.38	91.48	83.39
62.5	79.00	75.65	64.95	88.87	90.05
31.25	74.15	50.38	51.71	79.82	68.42
15.625	79.85	73.62	29.00	61.46	48.34
7.8125	48.59	42.39	17.49	46.63	46.73
$IC_{50} \\ (\mu g/mL)$	3.22	8.55	45.38	6.71	9.08

The study highlights the solvent-dependent biological activities of *Eucalyptus camaldulensis*. While methanol extracts were safer and richer in antioxidants, nonpolar fractions like n-hexane exhibited strong anti-inflammatory and cytotoxic effects. This suggests selective use of extracts depending on therapeutic goals. Further investigations, including isolation and structural elucidation of bioactive compounds and *in vivo* testing, are necessary to validate these *in vitro* findings

**Table 4:** Brine shrimp lethality test of *Eucalyptus camaldulensis* extracts

S/N	Sample	Conc. (µg/mL)	No. Of Brine shrimp	Replica	Mortality (24h)	% Mortality	LC50 (µg/mL)
1	Crude EtOH	1000	10	3	18	60	194.258
		100	10	3	13	43	
		10	10	3	11	37	
2	Methanol	1000	10	3	12	40	>1000
		100	10	3	10	33	
		10	10	3	9	30	
3	Ethyl acetate	1000	10	3	19	63	119.235
		100	10	3	15	50	
		10	10	3	10	33	
4	Chloroform	1000	10	3	26	87	3.970
		100	10	3	23	77	
		10	10	3	17	57	
5	n-Hexane	1000	10	3	29	97	3.810
		100	10	3	24	80	
		10	10	3	19	63	

# Conclusion

This study demonstrated that *Eucalyptus camaldulensis* leaf extracts possess solvent-dependent bioactivities. The methanol fraction showed the strongest antioxidant activity with no detectable cytotoxicity, indicating its safety and potential as a nutraceutical agent. Although the reduced cytotoxicity of the methanol fraction may imply a lower anticancer potential compared to the non-polar fractions, it also emphasizes its suitability for applications where safety and antioxidant efficacy are prioritized. In contrast, the n-hexane fraction exhibited potent anti-inflammatory and cytotoxic effects, suggesting the presence of bioactive lipophilic compounds with possible anticancer properties. These findings provide scientific justification for the ethnomedicinal use of *Eucalyptus camaldulensis* and highlight its potential as a source of natural therapeutic agents and for drug discovery. Further work should focus on isolation, structural characterization of active compounds, and *in vivo* validation.

#### **Conflict of interest**

The authors declare no conflict of interest

#### **Authors' Declaration**

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them

#### Acknowledgements

The authors gratefully acknowledge the Department of Pure and Industrial Chemistry, Bayero University, Kano, for providing laboratory facilities. Special thanks to the Herbarium Unit, Department of Biological Sciences, BUK, for authenticating the plant material. The assistance of laboratory staff, particularly Mr. Abubakar Rabil Khalil, for his technical guidance during the experimental procedures, is also sincerely appreciated.

# References

- Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. LWT Food Sci Technol. 1995; 28(1):25–30. <u>Doi: 10.1016/S0023-6438(95)80008-5</u>
- Kweki GR, Orhu A, Uzuegbu U, Iwhiwhu OS, Ohwokevwo OA. *In vitro* anti-inflammatory and antioxidant potentials of methanol extract of *Uvaria chamae* (bush banana) leaves. Trop J. Phytochem Pharm Sci. 2024; 3(1):85–92. Doi: 10.26538/tjpps/v3i1.6
- Mizushima Y, Kobayashi M. Interaction of antiinflammatory drugs with serum proteins. J. Pharmacol. 1968; 20(1):169–73. <u>Doi: 10.1111/j.2042-</u> 7158.1968.tb09718.x
- 4. Ogar VO, Itam YB, Akataobi US, Ukam EE, Ogar EA. *Effect* of ethanol and n-hexane combined extracts of selected plants on liver enzymes in Wistar rats. Trop J. Phytochem Pharm Sci. 2023; 2(4):65–72. Doi: 10.26538/tjpps/v2i4.4
- Ghareeb MA, Habib MR, Mossalem HS, Abdel-Aziz MS. Phytochemical analysis of *Eucalyptus camaldulensis* leaves extracts and testing its antimicrobial and schistosomicidal activities. Bull Natl Res Cent. 2018; 42(1):1–8. <u>Doi:</u> 10.1186/s42269-018-0017-2
- Uadia PO, Emmanuel CO, Oriakhi K, Imafidon KE. Anti-diabetic effect of *Eucalyptus camaldulensis (Red gum)* leaf-supplemented diet in streptozotocin-induced diabetic rats.
   Trop J. Pharm Res. 2024; 23(2):11. <u>Doi: 10.4314/tjpr.v23i2.11</u>

- Mohammed WH, Soud SA, Hameed AH, Hussein NN. Assessment of the cytotoxic activity of alcoholic extract of Eucalyptus camaldulensis on breast cancer cell line. Arch Razi Inst. 2021; 76(5):1307–1314. <u>Doi:</u> 10.22092/ari.2021.356104.1778
- Alabi MA, Abiola MM, Yakub NT, Ajani EO. In vitro antiinflammatory and anti-ulcerogenic potential of ethanol extract of *Carica papaya* leaves: phytochemical and molecular docking studies. Trop J. Phytochem Pharm Sci. 2025; 4(8):52–61. Doi: 10.26538/tjpps/v4i8.1/5
- Ayoola GA, Johnson OO, Aderounmu J, Raji L, Aremu RB, Bankole S. In vitro evaluation of the hypoglycaemic, antiinflammatory, and antioxidant activities of *Tragia benthamii Baker (Euphorbiaceae)*. Trop J. Phytochem Pharm Sci. 2024; 3(9):112–121. <u>Doi: 10.26538/tjpps/v3i9.4</u>
- Usman HN, Majiya H, Aremu KH, Legbo MI, Musa AD. Evaluation of phytochemical analysis and antibacterial potential of different solvent extracts of leaves and stem bark of *Eucalyptus camaldulensis* against selected pathogenic bacteria. *Biokemistri*. 2021; 33(4):301–310
- Gülcin I. Antioxidant activity of L-adrenaline: a structure– activity insight. Chem Biol Interact. 2009; 179:71–80. <u>Doi:</u> 10.1016/j.cbi.2008.09.023
- 12. Falodun A, Okafor OI, Erharuyi O, Okugbo OT. Phytochemical investigation and antioxidant activity evaluation of *Pyrenacantha staudtii (Icacinaceae) leaf.* Trop J. Phytochem Pharm Sci. 2025; 4(6):34–43. <u>Doi: 10.26538/tjpps/v4i6.2</u>
- Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, McLaughlin JL. Brine shrimp: a convenient general bioassay for active plant constituents. *Planta Med.* 1982; 45(5):31–4. DOI: 10.1055/s-2007-971236
- Amuamuta A. Cytotoxicity evaluation of *Eucalyptus camaldulensis leaf* extract against cholangiocarcinoma cell lines (CL-6). Res Sq. 2023. <u>Doi: 10.21203/rs.3.rs-2710703/v1</u>
- Nasr A, Khan TS, Zhu GP. Phenolic compounds and antioxidants from *Eucalyptus camaldulensis* as affected by some extraction conditions: a preparative optimization for GC–MS analysis. Prep Biochem Biotechnol. 2019; 49(5):1– 13. Doi: 10.1080/10826068.2019.1575860
- Siddique S, Iqbal M, Khan MA. Antioxidant activity of *Eucalyptus camaldulensis* extract. J. Pharm Anal. 2020; 10(3):237–244.
- Gautam PB, Kaur S, Gupta S. Phytochemical analysis of *Eucalyptus camaldulensis* leaves extract. J. Pharmacogn Phytochem. 2019; 8(1):2442–2446.
- Kumar P, Kumar V, Sharma S. Cytotoxic activity of *Eucalyptus camaldulensis* extract on breast cancer cells. J. Pharm Pharmacol. 2014; 66(8):1111–8.
- Nwaogu J, Argungu A, Adefarakan AJ. Antiulcer and antioxidant potential of *Eucalyptus camaldulensis* leaves methanol extract in albino rats. Asian J. Biochem Genet Mol Biol. 2021; 7(2):39–46. <u>Doi: 10.9734/ajbgmb/2021/v7i230172</u>
- El-Sayed AM, El-Ghany HM, Abd El-Hady MA. Nutritional evaluation of *Eucalyptus camaldulensis*. J. Anim Sci. 2017; 95(10):4331–4338.
- Ilboudo S, Valérie B, Jean E, Bazié T, Dado JN. Toxicological studies of *Eucalyptus camaldulensis* Dehnh (Myrtaceae) essential oil for the safety of its insecticidal application. Toxicol Risk Assess. 2024; 10(1): 1–8. Doi: 10.23937/2572-4061.1510061